

SPIFE® Split Beta SPE Procedure

Cat. No. 3399, 3398, 3399U, 3398U

INTENDED USE

The SPIFE Split Beta SPE System is intended for the separation of serum, urine, or cerebrospinal fluid (CSF) proteins by agarose gel electrophoresis using the SPIFE 3000 system.

For In Vitro Diagnostic Use.

RX Only

SUMMARY

Serum contains over one hundred individual proteins, each with a specific set of functions and subject to specific variation in concentration under different pathologic conditions.¹ Since the introduction of moving-boundary electrophoresis by Tiselius² and the subsequent use of zone electrophoresis, serum proteins have been fractionated on the basis of their electrical charge at a particular pH into five classical fractions: albumin, alpha₁, alpha₂, beta, and gamma proteins. Each of these classical electrophoretic zones, with the exception of albumin, normally contains two or more components. The relative proportions of these fractions have proven to be useful aids in the diagnosis and prognosis of certain disease states.³⁻⁵

PRINCIPLE

Proteins are large molecules composed of covalently linked amino acids. Depending on electron distributions resulting from covalent or ionic bonding of structural subgroups, proteins can be either polar or nonpolar at a given pH. In the SPIFE Serum Protein procedures, proteins are separated according to their respective electrical charges on agarose gel using both the electrophoretic and electroendosmotic forces present in the system. The proteins are then stained with a visible stain.

REAGENT

1. SPIFE Split Beta SPE Gel

Ingredients: Each gel contains agarose in a tris barbital/MOPS buffer with calcium lactate, a stabilizer, and a preservative.

WARNING: FOR IN-VITRO DIAGNOSTIC USE ONLY. The gel contains barbital which, in sufficient quantity, can be toxic.

Preparation for Use: The gels are ready for use as packaged.

Storage and Stability: The gels should be stored at room temperature (15 to 30°C) and are stable until the expiration date indicated on the package. The gels must be stored horizontally in the protective packaging in which they are shipped. **DO NOT REFRIGERATE OR FREEZE THE GELS.**

Signs of Deterioration: Any of the following conditions may indicate deterioration of the gel: (1) crystalline appearance indicating the agarose has been frozen, (2) cracking and peeling indicating drying of the agarose, (3) bacterial growth indicating contamination, (4) thinning of the gel blocks, (5) crystals in gel.

2. Acid Blue Stain

Ingredients: When dissolved as directed, the stain contains 0.5% (w/v) acid blue stain.

WARNING: FOR IN-VITRO DIAGNOSTIC USE ONLY. DO NOT INGEST.

Preparation for Use: Dissolve the dry stain (entire contents of vial) in 1 L of 5% acetic acid. Mix thoroughly for 30 minutes.

Storage and Stability: The dry stain should be stored at 15 to 30°C and is stable until the expiration date indicated on the package. The diluted stain is stable for six months when stored at 15 to 30°C.

Signs of Deterioration: The diluted stain should be a homogeneous mixture free of precipitate. Discard if precipitate forms.

3. Citric Acid Destain

Ingredients: After dissolution, the destain contains 0.3% (w/v) citric acid.

WARNING: FOR IN-VITRO DIAGNOSTIC USE. DO NOT INGEST - IRRITANT.

Preparation for Use: Pour 11 L of deionized water into the Destain vat. Add the entire package of Destain. Mix well until completely dissolved.

Storage and Stability: Store the Destain at 15 to 30°C. It is stable until the expiration date on the package.

Signs of Deterioration: Discard if solution becomes cloudy.

4. Acid Violet Stain (Optional Urine Stain)

Ingredients: The stain is comprised of Acid Violet stain.

WARNING: FOR IN-VITRO DIAGNOSTIC USE ONLY. DO NOT INGEST.

Preparation for Use: Dissolve the dry stain in 1 liter of 10% acetic acid and mix thoroughly. Fill the SPIFE stain vat.

Storage and Stability: The dry stain should be stored at 15 to 30°C and is stable until the expiration date indicated on the package. The stain solution is stable six months when stored at 15 to 30°C in a closed container.

Signs of Deterioration: The diluted stain should be a homogeneous mixture free of precipitate.

INSTRUMENT

A SPIFE 3000 Analyzer must be used to electrophorese, stain, destain, and then dry the gels. The gels may be scanned on a separate densitometer such as the QuickScan Touch Plus (Cat. No. 1640). Refer to the Operator's Manual for detailed instructions.

SPECIMEN COLLECTION AND HANDLING

Specimen: Fresh serum, urine, or CSF is the specimen of choice. Use of plasma will cause a fibrinogen band to appear as a distinct narrow band between the beta and gamma fractions.

Storage and Stability: If storage is necessary, serum samples may be stored covered at 15 to 30°C for 4 days, 2 to 8°C for 2 weeks, or -20°C for 6 months.⁶ Urine or CSF samples may be stored covered at 2 to 8°C for up to 72 hours or at -20°C for one month.

Urine Sample Preparation: Urine samples may be run diluted, neat, or concentrated. Shake samples to homogenize. Centrifuge desired volume at 2000 x g for 5 minutes. Remove supernatant and concentrate as follows:

| Total Protein (mg/dL) | Conc. Factor |
|-----------------------|--------------|
| < 50 | 100X |
| 50 - 100 | 50X |
| 100 - 300 | 25X |
| 300 - 600 | 10X |
| > 600 | 5X |

CSF Sample Preparation: CSF samples may be used after proper concentration (10 - 50X).

Interfering Factors:

- Hemolysis may cause false elevation in the alpha₂ and beta fractions.
- Inaccurate results may be obtained on specimens left uncovered, due to evaporation.

PROCEDURE

Materials provided: The kits can be ordered according to the matrix being tested. The applicator blade in the kit used for serum is different from the blade used for urine and/or CSF.

| Test Size | Cat. No. | |
|-------------|----------|-----------|
| | Serum | Urine/CSF |
| 100 Samples | 3398 | 3398U |
| 80 Samples | 3399 | 3399U |

The following materials needed for the procedure are contained in the SPIFE Split Beta SPE 80/100 Kits. Individual items are not available.

Cat. No. 3398, 3399

SPIFE Split Beta SPE Gels (10)
Acid Blue Stain (1 vial)
SPIFE Blotter C (10)
Citric Acid Destain (1 pkg)
Modified Applicator Blade Assembly-20 Sample

Cat. No. 3398U, 3399U

SPIFE Split Beta SPE Gels (10)
Acid Blue Stain (1 vial)
SPIFE Blotter C (10)
Citric Acid Destain (1 pkg)
SPIFE Applicator Blade Assembly-20 Sample

Material provided but not contained in the kit:

| ITEM | CAT. NO. |
|---|----------|
| SPIFE 3000 Analyzer | 1088 |
| QuickScan Touch Plus | 1640 |
| ESH Touch | 1380 |
| Applicator Blade Weights | 3387 |
| SPIFE Applicator Blades (for Urine and CSF) | 3450 |
| SPIFE Gel Block Remover | 1115 |

| | |
|---|--------|
| SPE Normal | 3424 |
| SPE Abnormal | 3425 |
| REP Prep | 3100 |
| SPIFE Dispo Sample Cups (deep well) | 3360 |
| SPIFE Dispo Sample Cups (for Urine & CSF) | 3369 |
| SPIFE 3000 20-100 Dispo Cup Tray | 3366 |
| SPIFE Urine/CSF Protein Accessory Kit | 3427 |
| SPIFE Urine IFE Alignment Tray | 3380 |
| Acid Violet Stain | 552351 |

Materials needed but not provided:

- 5% acetic acid
- 0.85% saline

STEP-BY-STEP METHOD

I. Sample Preparation

1. If testing 81 to 100 samples, remove five Disposable Applicator Blades from the packaging. If testing fewer samples, remove the appropriate number of Applicator Blades from the packaging. Remove the protective guard from the blades by gently bending the protective piece back and forth until it breaks free.



2. Place the five Applicator Blades into the numbered vertical slots 2, A, 9, 13, and 16 in the Applicator Assembly as given below. If using fewer Applicator Blades, place them into any of the slots noted above.

If testing only serum samples, follow the instructions marked “• Serum”. If testing serum with urine or CSF, follow instructions marked “• Serum and CSF or Urine”. Serum application is made after the second urine or CSF application. Therefore, the blade for serum application is not added until after the second urine/CSF application.

NOTE: The blade assembly will only fit into the slots in the Applicator Assembly one way; do not try to force the Applicator Blades into the slots.

3. Place an Applicator Weight on top of each Applicator Blade.
4. Slide the Disposable Sample Cup strips into the Cup Tray.
5. Pipette the following amount of sample into the cups. Cover the tray until ready to use.

NOTE: Application of Urine and CSF samples cannot be done with the Applicator Blades or cups packaged in the kit. Other blades (Cat. No. 3450) and cups (Cat. No. 3369) must be purchased.

| Sample | Volume | Blades | Cups |
|------------------|--------|--------|------|
| Serum or control | 45 µL | 3451 | 3360 |
| Urine or CSF | 20 µL | 3450 | 3369 |

Specimens with insufficient volumes may be run using the SPIFE Urine/CSF Accessory Kit (Cat. No. 3427) and the SPIFE Urine IFE Alignment Tray (3380).

If testing less than 81 samples, pipette samples into the rows of cups that correspond with Applicator Blade placement. Cover until ready to use.

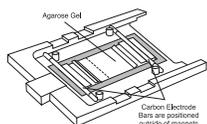
II. Gel Preparation

1. Remove the gel from the protective packaging and discard overlay.
2. Using a SPIFE Blotter C, gently blot the entire gel using slight fingertip pressure on the blotter. Remove the blotter.
3. Dispense approximately 2 mL of REP Prep onto the left side of the electrophoresis chamber.
4. Place the left edge of the gel over the REP Prep aligning the round hole on the left pin of the chamber. Gently lay the gel down on the REP Prep, starting from the left side and ending on the right side, fitting the obround hole over the right pin. Use lint-free tissue to wipe around the edges of the plastic gel backing, especially next to electrode posts, to remove excess REP Prep. Make sure no bubbles remain under the gel.
5. Clean the electrodes with deionized water before and after each use. Wipe with a lint-free tissue.
6. Place a carbon electrode on the outside ledge of each gel block outside the magnetic posts. Close the chamber lid.

III. Sample Application/Electrophoresis

Using the instructions provided in the appropriate Operator’s Manual, set up the parameters as follows for the SPIFE 3000.

Note: A “Dry 1” time of 10 or 12 minutes is



recommended. However, due to variations in environmental conditions, the following ranges are acceptable.

*Dry 1 = 10-15 minutes.

• Serum - Choose one of two application options

• Option 1

Electrophoresis Unit

- 1) No Prompt
Load Sample 1 00:30 21°C SPD1
- 2) No Prompt
Apply Sample 1 00:30 21°C SPD1 LOC1
- 3) No Prompt
Electrophoresis 1 6:00 21°C 650V 130 mA
- 4) Remove gel blocks, (continue)
Dry 1 *10:00 54°C
- 5) No prompt
END OF TEST

• Option 2

- 1) No Prompt
Load Sample 1 00:30 21°C SPD1
- 2) No Prompt
Apply Sample 1 00:10 21°C SPD1 LOC1
- 3) No Prompt
Absorb 1 00:45 21°C
- 4) No Prompt
Electrophoresis 1 6:00 21°C 650V 130 mA
- 5) Remove gel blocks, (continue)
Dry 1 *10:00 54°C
- 6) No prompt
END OF TEST

• Serum and CSF or Urine

Electrophoresis Unit

- 1) No Prompt
Load Sample 1 00:30 21°C SPD1
- 2) No Prompt
Apply Sample 1 00:30 21°C SPD1 LOC1
- 3) No Prompt
Load Sample 2 00:30 21°C SPD1
- 4) No Prompt
Apply Sample 2 00:30 21°C SPD1 LOC1
- 5) To Continue, (continue)
Load Sample 3 00:30 21°C SPD1
- 6) No Prompt
Apply Sample 3 1:00 21°C SPD1 LOC1
- 7) No Prompt
Absorb 1 1:00 21°C
- 8) No Prompt
Electrophorese 1 6:00 21°C 650V 130 mA
- 9) Remove gel blocks, (continue)
Dry 1 *10:00 54°C
- 10) No Prompt
END OF TEST

Stainer Unit

• Serum and CSF or Urine

NOTE: If testing urine samples with Acid Violet Stain, change "VALVE = 3" to "VALVE = 5" in Step 1.

- 1) No Prompt
Stain 1 4:00 REC = OFF VALVE = 3
- 2) No Prompt
Destain 1 1:00 REC = ON VALVE = 2
- 3) No Prompt
Destain 2 1:00 REC = ON VALVE = 2
- 4) No Prompt
Destain 3 1:00 REC = ON VALVE = 2
- 5) No Prompt
Dry 1 *12:00 63°C
- 6) No Prompt
END OF TEST

1. Press the **TEST SELECT/CONTINUE** button located on the electrophoresis side of the instrument until the **SPLIT BETA SPE or URINE PROTEIN** option appears on the display. Open the chamber lid.

- Place the Cup Tray with samples on the SPIFE 3000. Align the holes in the tray with the pins on the instrument. Close the chamber lid.
- With the appropriate display, press the **START/STOP** button. An option to either begin the test or skip the operation will be presented. Press **START/STOP** again to begin. If testing serum only, the SPIFE 3000 will apply the samples, electrophorese, and beep when completed.
- If testing serum and urine/CSF, open the chamber lid after the beep. Place the Modified Blade in the Applicator Assembly for serum application. Close the chamber lid. Press **TEST SELECT/CONTINUE**.

IV. Visualization

- After electrophoresis is complete, open the chamber lid and use the SPIFE Gel Block Remover to remove the gel blocks. Replace the electrodes on each end of the gel to prevent curling during drying.
- Close the chamber lid and press the **TEST SELECT/CONTINUE** button to dry the gel.
- After the gel has been dried, carefully remove the gel from the electrophoresis chamber.
- Remove the Gel Holder from the stainer chamber. Attach the gel to the holder by placing the round hole in the gel backing over the left pin on the holder and the obround hole over the right pin on the holder.
- Place the Gel Holder with the attached gel facing backwards into the stainer chamber.
- With the appropriate test name on the display, press the **START/STOP** button. An option to either begin the test or skip the operation will be presented. Press **START/STOP** to begin. The instrument will stain, destain, and dry the gel.
- When the process is completed, the instrument will beep. Remove the Gel Holder from the stainer and scan the bands in a densitometer.

Evaluation of the Protein Bands

Qualitative evaluation: The urine and CSF samples run on the SPIFE Split Beta SPE Gel can only be qualitatively inspected for the presence of the bands.

Quantitative Evaluation: Scan the serum samples on the SPIFE Split Beta SPE Gel in the QuickScan Touch Plus agarose side up on the acid blue setting. A slit size of 5 is recommended.

Stability of End Product: The completed, dried SPIFE Split Beta SPE Gel is stable for an indefinite period of time.

Quality Control: SPE Normal (Cat. No. 3424) and SPE Abnormal (Cat. No. 3425) may be used to verify all phases of the procedure and should be used on each gel run. If desired, a control or patient sample may be diluted 1:7 with 0.85% saline (1 part sample + 6 parts saline) and run with urines and CSFs for qualitative comparison. Refer to the package insert provided with the control for assay values.

REFERENCE VALUES

The reference range presented was established with the SPIFE Split Beta SPE and the QuickGel Split Beta SPE Systems on 48 normal specimens using the SPIFE 3000 Analyzer. These values are presented as a guideline.

| <u>Protein Fraction</u> | <u>% of Total Protein</u> <u>X ± 2 S.D.</u> |
|-------------------------|--|
| Albumin | 47.6 - 61.9 |
| Alpha ₁ | 1.4 - 4.6 |
| Alpha ₂ | 7.3 - 13.9 |
| Beta | 10.9 - 19.1 |
| Gamma | 9.5 - 24.8 |

Each laboratory should perform its own normal range study.

Variations of Expected Values⁵

Studies show that values are the same for both males and nonpregnant females. (Some differences are seen in pregnant females at term and in women on oral contraceptives.) Age has some effect on normal levels. Cord blood has decreased total protein, albumin, alpha₂, and beta fractions, slightly increased alpha₁, and normal or increased gamma fractions (largely of maternal origin). The gamma globulins drop rapidly until about three months of age, while the other fractions have reached adult levels by this time. Adult levels of the gamma globulins are not reached until 16 years of age. The albumin decreases and beta globulin increases after the age of 40.

Further Testing Required

The serum protein electropherogram or densitometric tracing should be evaluated for abnormalities. If abnormalities are observed, appropriate follow-up studies should be initiated. These may include immunoelectrophoresis, immunofixation, quantitation of immunoglobulins, bone marrow examination, and other appropriate tests.

RESULTS

Figure 1 illustrates the electrophoretic mobilities of the albumin, alpha₁, alpha₂, beta, and gamma protein bands on SPIFE Split Beta SPE-100 Gel. The fastest moving band, and normally the most prominent, is the albumin band found closest to the anodic edge of the gel. The faint band next to this is alpha₁, followed by alpha₂, globulin, beta, and gamma globulins.



Figure 1: A SPIFE Split Beta SPE-100 Gel showing relative position of the bands.

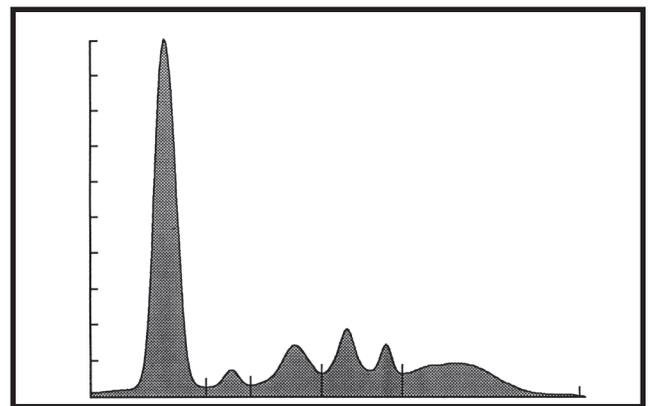


Figure 2: A scan of a SPIFE Split Beta SPE pattern.

Calculations of the Unknown

The Helena QuickScan Touch Plus densitometer will automatically calculate and print the relative percent and the absolute value of each band when the total protein is entered. Refer to the Operator's Manual provided with the instrument.

INTERPRETATION OF RESULTS⁶

Results on normal individuals will cover age and sex-related variations and day-to-day biologic variations. Disease states in which abnormal patterns are observed include inflammatory response, rheumatic disease, liver diseases, protein-loss disorders, plasmacell dyscrasias, pregnancy, and genetic deficiencies.

LIMITATIONS

Since all electrophoretic procedures are nonlinear, it is critical to fill the wells with the recommended volume of undiluted serum to obtain optimal resolution and reproducible results. Noncompliance with the recommended procedure may affect the results.

SPECIFIC PERFORMANCE CHARACTERISTICS

PRECISION

Within Run: A normal and an abnormal control were run alternately 50 times each on a single gel with the following results: N = 50

| <u>Protein Fraction</u> | <u>Mean %</u> | <u>SD</u> | <u>CV</u> |
|-------------------------|---------------|-----------|-----------|
| Albumin | 57.7 | 2.4 | 4.2% |
| Alpha ₁ | 3.2 | 0.6 | 17.6% |
| Alpha ₂ | 11.1 | 0.7 | 6.8% |
| Beta | 15.1 | 0.8 | 5.4% |
| Gamma | 12.9 | 1.1 | 8.7% |

Abnormal Control

| <u>Protein Fraction</u> | <u>Mean %</u> | <u>SD</u> | <u>CV</u> |
|-------------------------|---------------|-----------|-----------|
| Albumin | 47.3 | 1.4 | 3.0% |
| Alpha ₁ | 3.1 | 0.4 | 11.4% |
| Alpha ₂ | 9.5 | 0.5 | 4.8% |
| Beta | 11.6 | 0.6 | 4.8% |
| Gamma | 28.5 | 0.6 | 2.0% |

Between-Run: A normal and an abnormal control were run alternately 50 times each on three gels with the following results: N = 150

Normal Control

| <u>Protein Fraction</u> | <u>Mean %</u> | <u>SD</u> | <u>CV</u> |
|-------------------------|---------------|-----------|-----------|
| Albumin | 58.9 | 2.7 | 4.6% |
| Alpha ₁ | 3.0 | 0.6 | 21.0% |
| Alpha ₂ | 11.0 | 0.7 | 6.4% |
| Beta | 15.1 | 0.9 | 6.3% |
| Gamma | 12.1 | 1.3 | 10.6% |

Abnormal Control

| <u>Protein Fraction</u> | <u>Mean %</u> | <u>SD</u> | <u>CV</u> |
|-------------------------|---------------|-----------|-----------|
| Albumin | 47.9 | 1.6 | 3.3% |
| Alpha ₁ | 3.0 | 0.4 | 14.4% |
| Alpha ₂ | 9.4 | 0.5 | 5.3% |
| Beta | 11.5 | 0.5 | 5.7% |
| Gamma | 28.2 | 0.6 | 2.1% |

CORRELATION

Normal (N = 36) and abnormal (N = 43) serum samples were analyzed using the SPIFE SPE Vis-60 system and the SPIFE Split Beta SPE system.

N = 79

Y = 1.02X - 0.31

R = 0.99

X = SPIFE SPE Vis-60

Y = SPIFE Split Beta SPE

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